

Supplemental Material

Myeloid DRP1 Deficiency Limits Revascularization in Ischemic Muscles via Inflammatory Macrophage Polarization and Metabolic Reprogramming

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Running Title: Macrophage Drp1 and angiogenesis

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Supplemental Figure S1

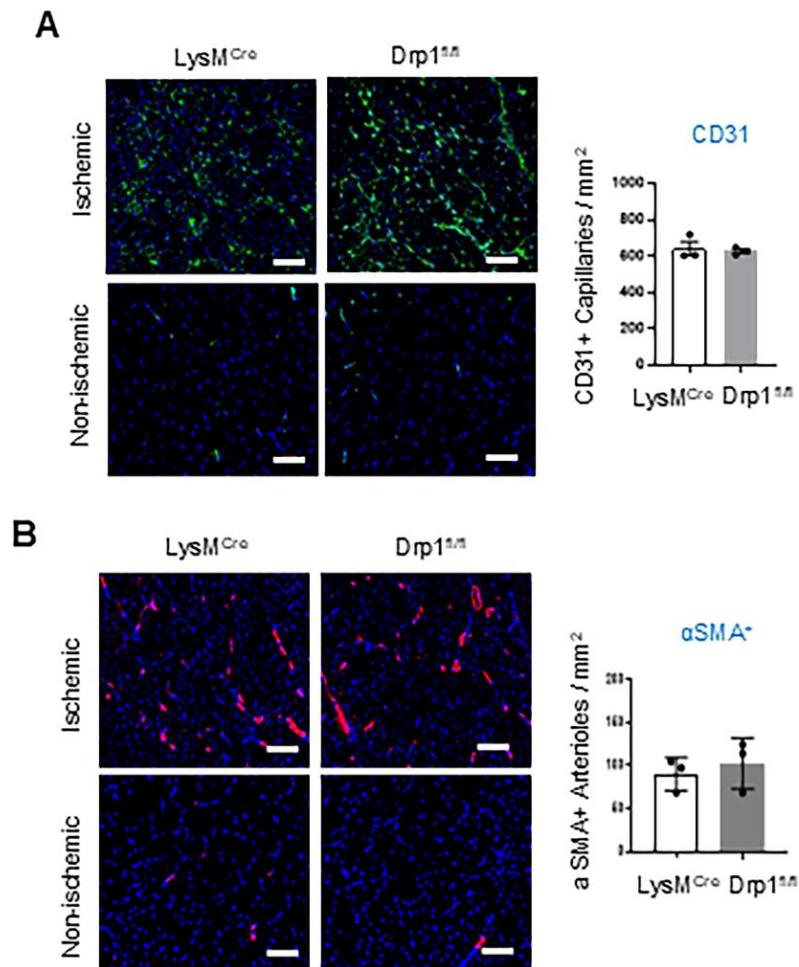


Figure S1: *Drp1^{fl/fl}* and *LysM-Cre* mice exhibited no difference in the number of CD31⁺ capillary endothelial cell (EC) and α SMA⁺ arterioles in ischemic muscles following hindlimb ischemia (HLI). A and B. Immunofluorescence (IF) images (left) and quantification (right) of CD31⁺ capillary-like EC (A) and α -smooth muscle cells (SMA⁺) (arterioles) (B) in ischemic and non-ischemic GC muscles at day 21 post-HLI. scale bar=20 μ m. n=3. Data are mean $\pm$ SEM.

Supplemental Figure S2

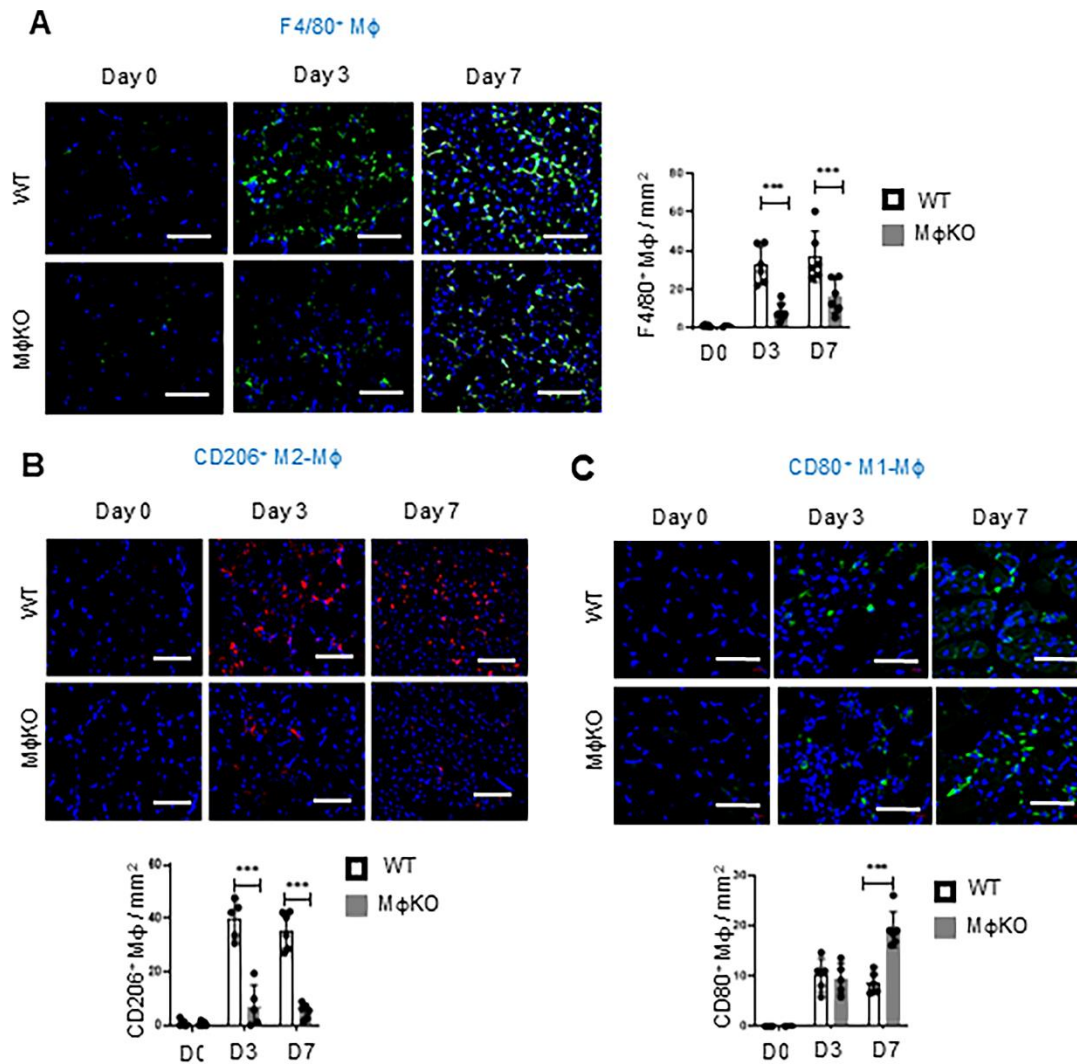


Figure S2: Myeloid *Drp1*^{KO} mice exhibited a reduction in F4/80⁺ macrophages and anti-inflammatory CD206⁺ M2-like macrophages, along with an increase in pro-inflammatory CD80⁺ M1-like macrophages in ischemic muscles. **A.** IF analysis of F4/80⁺ macrophages (green) with DAPI (blue) staining in non-ischemic and ischemic GC muscles of WT and M ϕ KO mice at the indicated times after HLI. Scale bar=20 μ m. **B-C** IF analysis of CD206⁺ (red) M2-like macrophages (**B**) and CD80⁺ (green) M1-like macrophages (**C**) with DAPI (blue) in ischemic GC muscles of WT and M ϕ KO mice at the indicated times post-HLI. Scale bar=20 μ m, bottom panels show quantification. n=3-5 mice per group, (two-way ANOVA followed by Tukey's multiple comparison test). Data are mean $\pm$ SEM. ***p<0.001.

Supplemental Figure S3

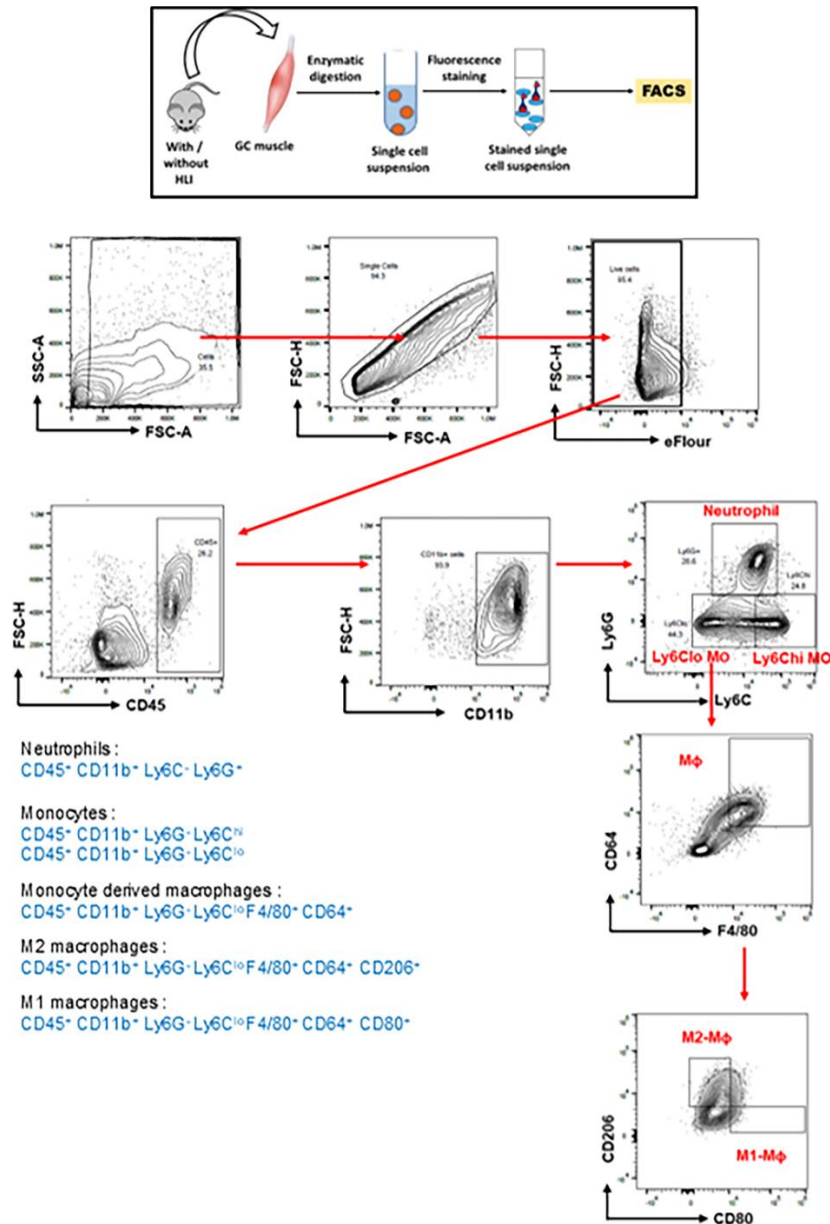


Figure S3: Schematic representation of the gating strategy for flowcytometry based immunophenotyping of gastrocnemius muscle. Gating strategy for flow cytometry-based analysis to identify and quantify neutrophils, monocytes, naïve (M0) macrophages, pro-inflammatory M1 macrophages, and anti-inflammatory M2 macrophages isolated from ischemic GC muscle at the indicated time after HLI. Cells were first gated based on FSC-A and SSC-A for singlets, followed by gating for live cells and CD45⁺/CD11b⁺ double-positive leukocytes. Neutrophils (Ly6G⁺), monocyte (Ly6C^{hi}), naïve M0 Macrophages (Ly6C^{lo}/F4/80⁺/CD64⁺), M1 macrophages (Ly6C^{lo}/F4/80⁺/CD64⁺/CD80⁺), and M2 macrophages (Ly6C^{lo}/F4/80⁺/CD64⁺/CD206⁺) were quantified. Note CD80⁺CD206⁺ double positive MΦ in M1/M2 contour plots are not considered in the quantification.

Supplemental Figure S4

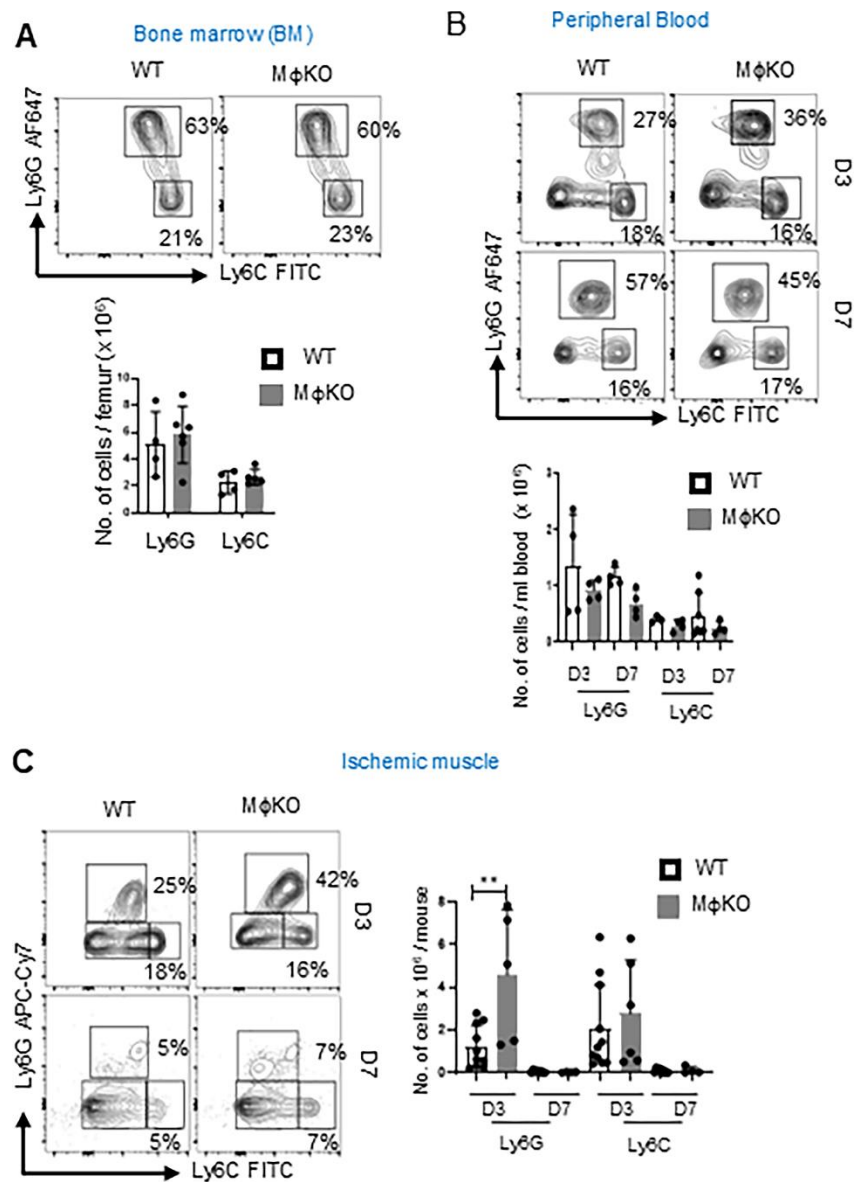


Figure S4: Neutrophils and monocytes in peripheral blood, bone marrow (BM), and ischemic GC muscles of WT and Myeloid *Drp1*^{KO} mice following HLI. Representative flow cytometry contour plots and quantification of Ly6G^{hi} neutrophils and Ly6C^{hi} monocytes in the BM (A) and peripheral blood (B) at day 3 post-HLI, as well as in ischemic GC muscle at days 3 and 7 post-HLI in WT and M ϕ KO mice. n=3-10 mice per group. Data are mean $\pm$ SEM. n=3-5. **p<0.01.

Supplemental Figure S5

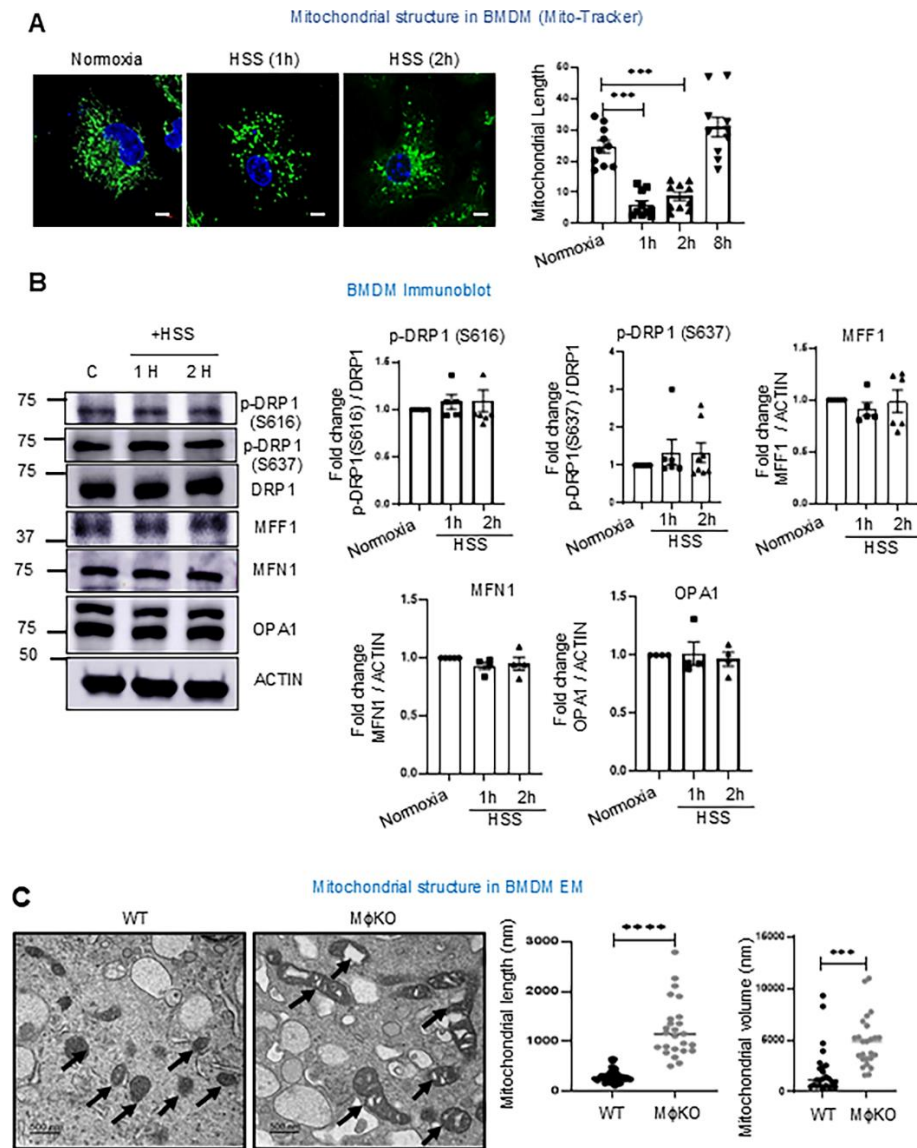


Figure S5: BMDMs under HSS exhibited increased mitochondrial fission without changes in the expression of key mitochondrial fission/ fusion proteins, while *Drp1*^{KO} BMDMs under HSS showed mitochondrial fusion. **A.** Mitochondrial structure in WT-BMDMs stained with MitoTracker Green after HSS stimulation for the indicated time (left). The right panel shows quantification of mitochondrial length measured using image J (right). Scale bar=5μm, n=number of cells analyzed from 3 independent exp. ***p<0.001 (one-way ANOVA followed by Tukey's multiple comparison test). **B.** Western blot analysis of p-DRP1-S616, p-DRP1-S637, total DRP1, MFF1, MFN1, OPA1 and ACTIN (loading control) in WT-BMDM exposed to HSS for the indicated time. The right panels show quantification. Data are mean ± SEM. n=4-5. **C.** Electron microscopy images and quantification of mitochondrial length and volume in WT and MφKO BMDMs after 1 h of HSS stimulation. n=number of cells analyzed per group. Data are mean ± SEM. ****p<0.0001 (unpaired t test).

Supplemental Figure S6

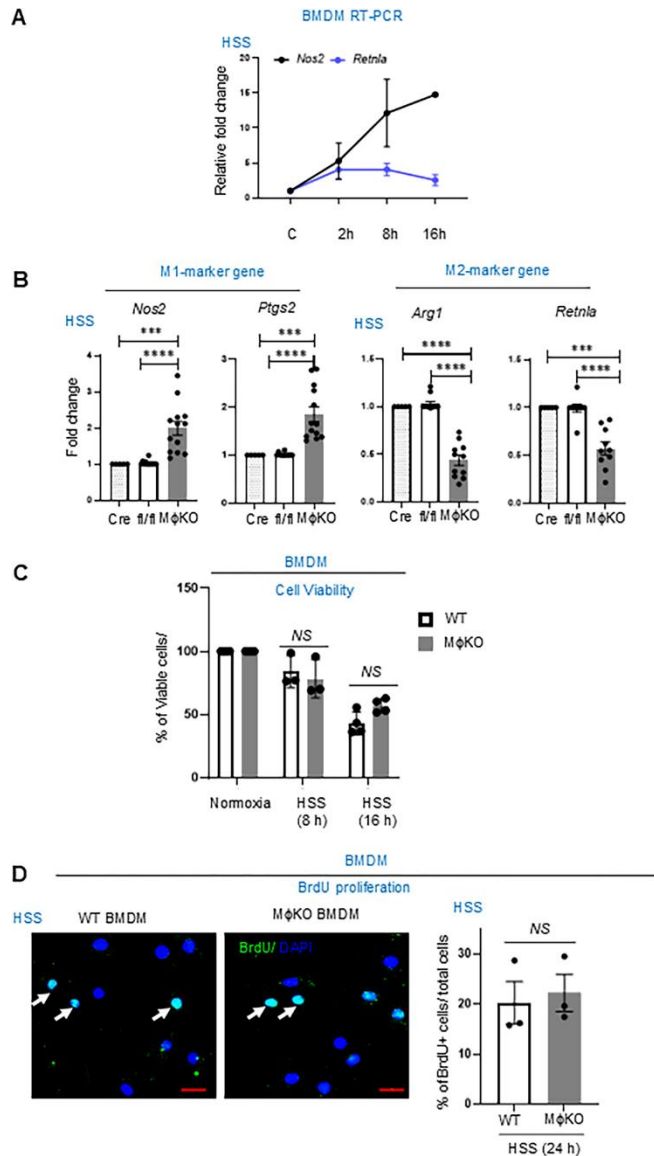


Figure S6: *Drp1*^{KO} BMDMs under HSS showed an increase in M1-marker genes and a decrease in M2-marker gene expression without affecting cell viability or proliferation. **A. Quantitative RT-PCR (relative to *Hprt*) analysis of M1 marker gene *Nos2* and M2 marker gene *Retnla* mRNA levels in WT-BMDMs exposed to HSS for the indicated time. Data are expressed as fold change from normoxic controls. **B.** mRNA levels of M1 markers (*Nos2* and *Ptgs2*) and M2 markers (*Arg1* and *Retnla*) in *LysM*-Cre WT, *Drp1*^{fl/fl} WT, and MφKO BMDMs under HSS for 8 h. Data are mean ± SEM. n=3-7. ****p<0.0001, ***p<0.001 (one-way ANOVA followed by Tukey's multiple comparison test). **C.** Cell viability in WT and MφKO BMDMs exposed to either HSS or normoxia for the indicated time, as measured by a CCK-8-based colorimetric assay. **D.** IF images (left) and quantification (right) of proliferating BrdU⁺ (green) with nuclei stained by DAPI (blue) after 24 h of HSS exposure in WT and MφKO BMDMs. Scale bar=20 μm. Data represent n=3-4 independent experiments and are shown as mean ± SEM.**

Supplemental Figure S7

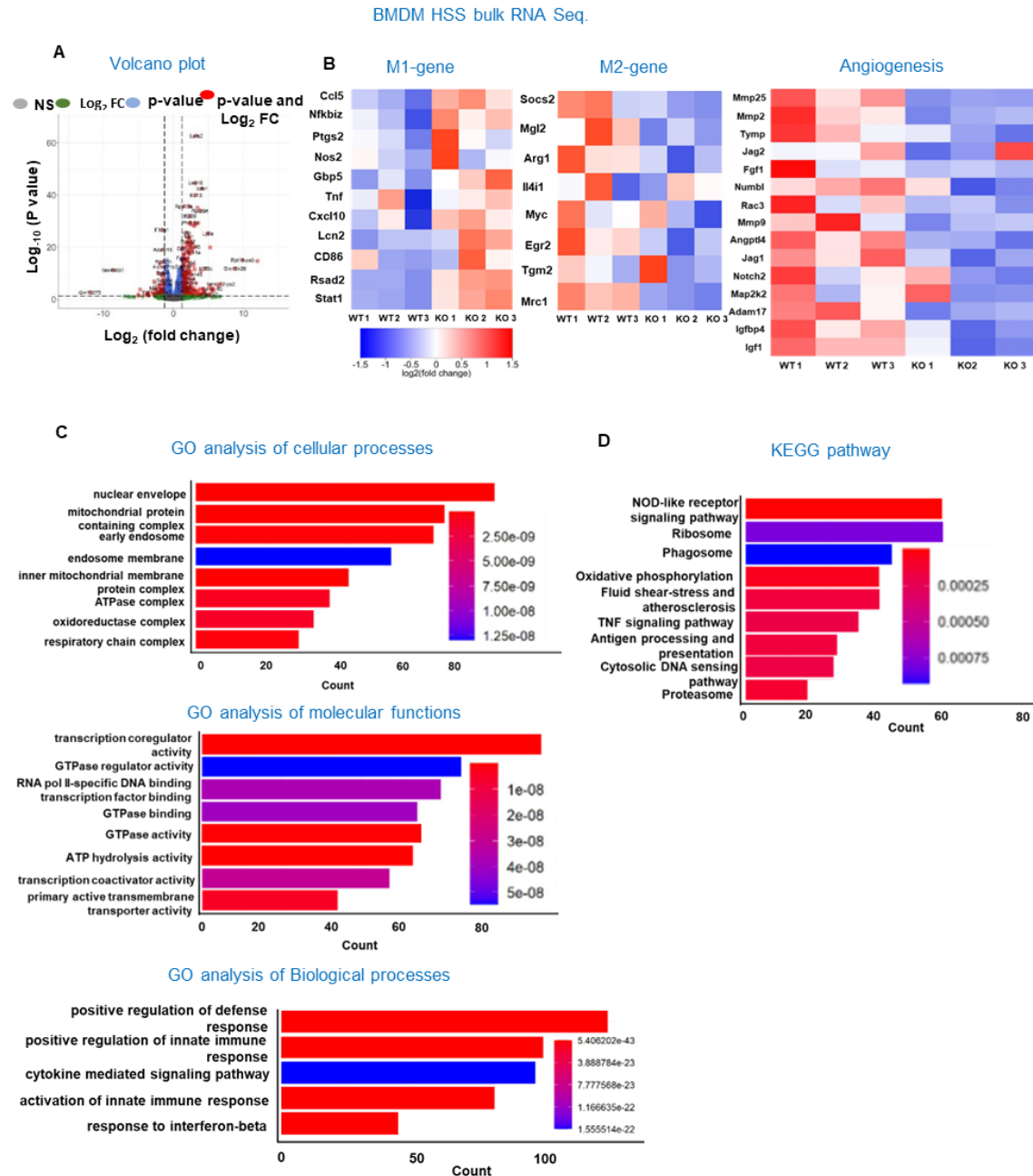


Figure S7: *Drp1*^{KO} BMDM exposed to HSS exhibited increased expression of pro-inflammatory M1 genes and decreased anti-inflammatory M2 genes and pro-angiogenic genes. RNA-Seq analysis was performed on BMDMs from WT and M ϕ KO mice (n = 3 per group) that were cultured for 7 days and then exposed to HSS for 8h. **A.** A volcano plot showing the differentially expressed genes. **B.** Heat maps depicting the expression patterns of pro-inflammatory M1 genes, anti-inflammatory M2 genes, and pro-angiogenic genes. **C.** Gene Ontology (GO) enrichment analysis of the differentially expressed genes. **D.** KEGG pathway enrichment analysis of the differentially expressed genes.

Supplemental Figure S8

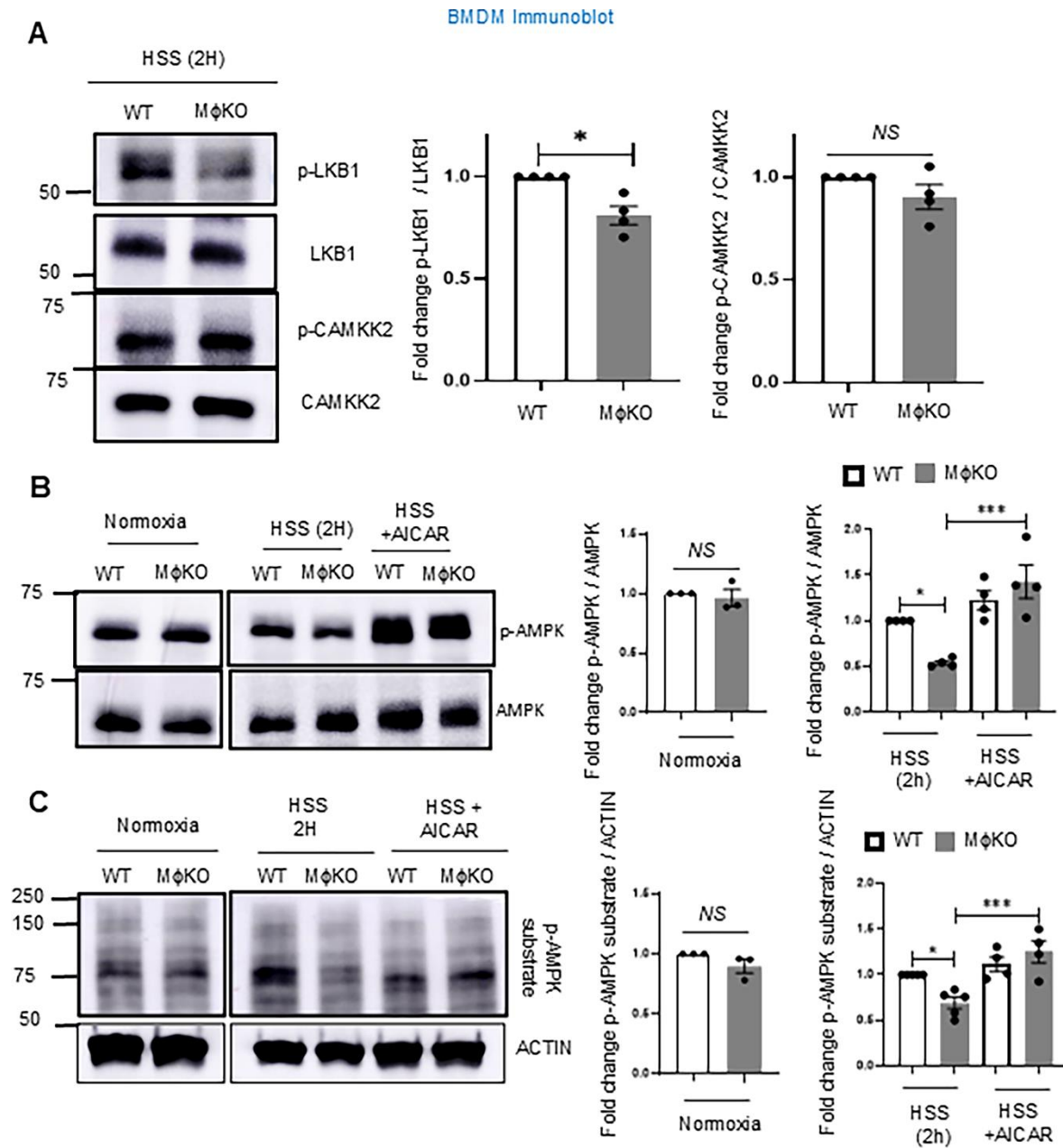


Figure S8: Reduced AMPK activation in *Drp1*^{KO} BMDM under HSS was restored by AMPK activator AICAR. WT and M ϕ KO BMDMs were subjected to 2 h of HSS stimulation and analyzed by Western blot using anti- p-LKB1, LKB1, p-CAMKK2 and CAMKK2 antibodies. Right panels show quantification. Data are mean $\pm$ SEM. (n=4, unpaired t test). **B.** WT and M ϕ KO BMDMs were exposed to normoxia or HSS for 2 h with or without AICAR pre-treatment (100 μ M for 2h) and analyzed by Western blot using anti-p-AMPK, AMPK, total p-AMPK substrate, and ACTIN antibodies. Right panels show quantification. Data are mean $\pm$ SEM. n=4-5. (ANOVA followed by Tukey's multiple comparison test).

Supplemental Figure S9

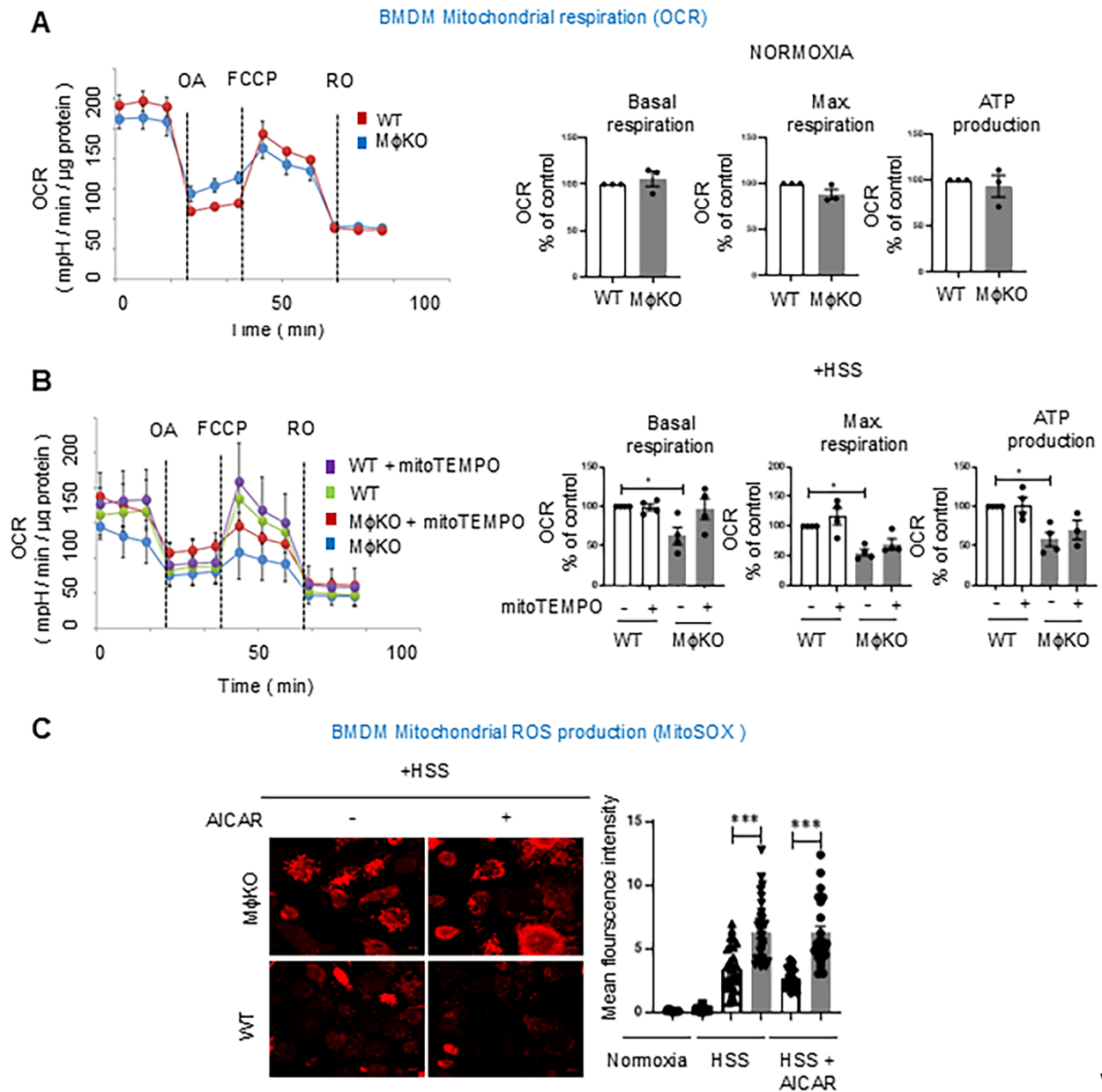


Figure S9: Excess mitoROS production in *Drp1*^{KO} BMDMs under HSS does not precede mitochondrial dysfunction and is not suppressed by the AMPK activator. **A.** Mitochondrial respiration (OCR) measured via Seahorse XF analyzer in WT and MφKO BMDMs under normoxia for 2h. **B.** OCR in WT and MφKO BMDMs after 2h of HSS stimulation, with or without mitoTEMPO pre-treatment (16 h at 20 μM). Right panels show quantification. Data represent n=3 (one-way ANOVA followed by Tukey's multiple comparison test). **C.** Left, Mitochondrial ROS production measured by MitoSOX after 2 h of HSS stimulation in WT and MφKO BMDMs, with or without AMPK activator AICAR pre-treatment (2 h at 100 μM). Right: Quantification of mean fluorescence intensity using ImageJ. Scale bars=5μm. *p<0.05, ***p<0.001 (one-way ANOVA followed by Tukey's multiple comparison test). Data are mean ± SEM.